NEUROMICS

GFP-Immortalized Human Brain Microglia Cells

Catalog #: HBMCs001-GFP

Cell #: >5x10^5 cells

Storage: Liquid Nitrogen until ready for culture.
While Culturing keep in 37°C CO_2 incubator

Product Format: Frozen Vial

GENERAL INFORMATION

Human brain microglia cells are isolated from healthy human brain tissue. GFP-Immortalized Human Brain Microglia Cells are selected from HBMgs infected with lentiviruses expressing hTert and GFP with puromycin. HBMgs can be cultured in long-term (tested >20 passages). It is recommended to use Alpha-glia Expansion medium (AGEM-001) for the culturing of HBMgs

Product is for Research use only.

Frozen Vials are shipped in a Dry Ice Package.

CHARACTERIZATION OF THE CELLS

Cytoplasmic F4/80 Positive

CD68 Positive

HBMgs are tested negative for HIV-1, HBV, HCV, and mycoplasma.

HANDLING OF ARRIVING CELLS

When you receive the cells in a frozen vial, you can transfer the vial of cells into a -80C freezer for short period storage or a liquid nitrogen tank for long-term storage. Thaw the cells in a 37C water bath, and then transfer the cells into a T25 flask pre-coated with Universal Coating Solution (AC002) as described in following details.

PROTOCOL FOR THAWING THE CELLS AND SUBCULTURE

1. Prepare a flask (T-25 flask is recommended) by adding 5 ml of Universal Coating Solution (AC002) to a T-25 flask, leave the flask in an incubator overnight (minimum one hour in 37C incubator).

2. Rinse the coated flask with sterile PBS twice and add 7 ml of Alpha-glia Expansion medium (AGEM-001) to the flask. Leave the flask in the hood and go to thaw the cells.

3. Warm Alpha-glia Expansion medium before thawing the cells.

4. Place the vial in a 37C water bath, hold and rotate the vial gently until the contents are completely thawed. Remove the vial from the water bath immediately, wipe it dry, rinse the vial with 70% ethanol and transfer it to a sterile field. Remove the cap, being careful not to touch the interior threads with fingers. Using 1 ml Eppendorf pipette gently resuspend the contents of the vial

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5. Dispense the contents of the vial into the neutralized coated culture vessels. A seeding density of more than 10,000 cells/cm2 is recommended.

6. Replace the cap or cover, and gently rock the vessel to distribute the cells evenly. Loosen cap if necessary, to permit gas exchange.

7. Return the culture vessels to the incubator (For best result, do not disturb the culture for at least 16 hours after the culture has been initiated. Change the growth medium the next day to remove the residual DMSO by centrifuging the unattached cells down, resuspending the cells with 5ml fresh medium, and add the cells back to the flask. Then change the medium every other day thereafter).

8. Change the medium every two to three days thereafter.

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