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PCR DNA extraction and purification magnetic beads kit Data Sheet

Catalog Number:	EP10013	**Size:**	100T
Kit Components included:	• Si-Mag magnetic beads – 5 ml • PCR DNA binding buffer – 15 ml • Elution Buffer – 4 ml		
Materials needed but not provided with the kit:	• 80% Ethanol • Si-Mag Magnet (sold separately)		
Applications:	This kit provides a simple, rapid and efficient method for the recovery and purification of DNA directly from Agarose gel (100 bp to 50 kb) with typical recovery efficiency up to 85%.		
Storage:	Magnetic beads should be stored at 2-8°C, but other kit reagents need to be stored at room temperature. Avoid repeated freeze-thaw cycles.		

Introduction

This kit provides a simple, rapid and efficient method for the recovery and purification of DNA directly from PCR products (100 bp to 50 kb) with typical recovery efficiency up to 85%. The resulting product can be directly used for sequencing, restriction digestion, or PCR and other downstream experiments. In addition, the kit can be used to concentrate DNA.
The kit will work with a 96 well round bottom plates if a special magnetic frame is used.
The kit can also be used with a variety of automatic nucleic acid extraction instruments and workstation.

Precautions

1. Avoid freeze/thaw cycles and centrifugation which could damage the beads.
2. Be sure to mix well before using magnetic beads, can be vortexed about 10 seconds.
3. Vortex samples for about 10 seconds before adding
4. Elute DNA from the beads completely.

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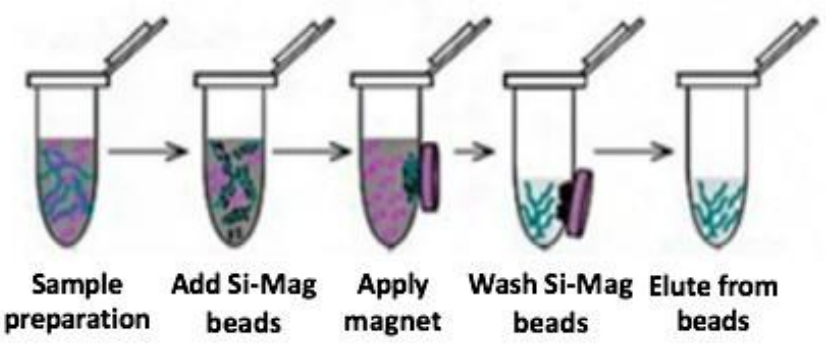
Principle of Assay:



Procedure for purification of PCR DNA product:

1. **Sample preparation.** Add **3 volumes** of PCR DNA binding solution directly to the PCR product. For example, add 120 ul of PCR DNA binding buffer to a 40 ul of PCR product.
2. **Transfer** all content to an Eppendorf tube, then add **50 ul** of magnetic beads, mix well and incubate 3-5 min at RT . Put Eppendorf tube onto the Si-Mag magnet rack for 20 seconds. Remove supernatant by holding the magnet rack upside or by pipetting.
3. **Wash** the beads with **500µL** of 80% ethanol **twice.**
4. **Dry** the beads at 55°C for 8 min leaving the tube open. **Do not over-dry the beads.**
5. **Elute** the DNA from beads with **35 ul** of elution buffer, incubate for at least 2 min and then vortex at full speed for 1 min. **Alternatively,** incubation at 60°C for 2 min may improve the recovery for DNA larger than 3 kb.
6. **Remove beads** by using magnet rack, pipette DNA out and transfer to a clean tube.
7. **Store purified** DNA at -20°C for long-term storage.

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