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Agarose Gel DNA extraction and purification magnetic beads kit Data Sheet

Catalog Number:	EP10010	**Size:**	100T

Kit Components included:

- Si-Mag magnetic beads 5ml
- Gel solvent buffer 50 ml . **For purification of DNA from Agarose gel**
- Wash Solution 25 ml (add 25 mL of Isopropanol)
- Elution Buffer 5 ml

Materials needed but not provided with the kit:

- 80% Ethanol
- Si-Mag Magnet (sold separately)
- Isopropanol (ACS grade)

Applications: This kit provides a simple, rapid and efficient method for the recovery and purification of DNA directly from Agarose gel (100 bp to 50 kb) with typical recovery efficiency up to 85%.

Storage: Magnetic beads should be stored at 2-8°C but other kit reagents need to be stored at room temperature.

Introduction

This kit provides a simple, rapid and efficient method for the recovery and purification of DNA directly from Agarose gel (100 bp to 50 kb) with typical recovery efficiency up to 85%. The resulting product can be directly used for sequencing, restriction digestion, or PCR and other downstream experiments. In addition, the kit can be used to concentrate DNA.

The kit will work with a 96 well round bottom plates if a special magnetic frame is used. The kit can also be used with a variety of automatic nucleic acid extraction instruments and workstation.

Precautions

1. Avoid freeze/thaw cycles and centrifugation which could damage the beads.
2. Be sure to mix well before using magnetic beads, can be vortexed about 10 seconds.
3. Vortex samples for about 10 seconds before adding
4. Elute DNA from the beads completely.

Neuromics Antibodies • 5325 West 74th Street, Suite 8 • Edina, MN 55439
phone 866-350-1500 • fax 612-677-3976 • e-mail pshuster@neuromics.com

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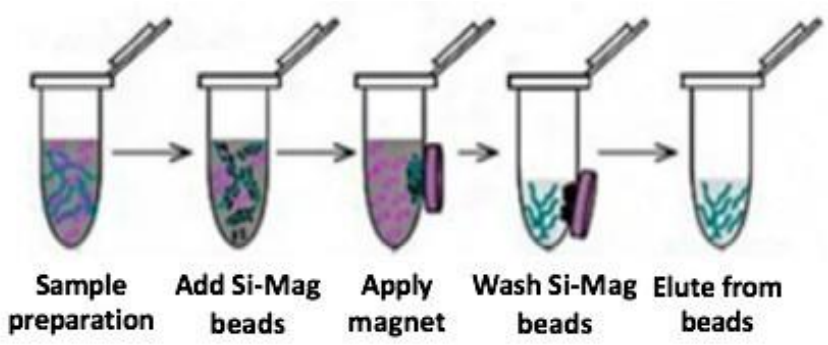
Principle of Assay:

Procedure for purification of DNA from agarose gel:

1. **Sample preparation.** Add **500 ul** of gel solvent buffer and a gel slice of up to **400mg** into a clean Eppendorf tube. For a gel slice larger than 400mg, add at least 800 ul of gel solvent buffer. Incubate at **65°C** for 10 min or until the gel is completely dissolved. Vortex the tube periodically to ensure complete dissolving.
2. **Transfer** all content to an Eppendorf tube and then add **50 ul** of magnetic beads, mix well and incubate 3-5 min at RT. Put Eppendorf tube onto the Si-Mag magnet rack for 20 seconds. NOTE: The estimated recovery of DNA is bout 2 ug per 50 ul of beads
3. **Remove** supernatant by holding the magnet rack upside down or by pipetting.
4. **Wash** the beads with **500 ul** of Wash Solution. *Make sure the beads get completely resuspended by vortexing and then repeat step 3.*
5. **Wash** the beads with **500µL** of 80% ethanol.
6. **Make Sure** the beads get completely resuspended by vortexing and the repeating step 3.
7. **Dry** the beads at 55°C for 8 min leaving the tube open. **Do not over-dry the beads.**
8. **Elute** the DNA from beads with **35 ul** of elution buffer, incubate for at least 2 min and then vortex at full speed for 1 min. **Alternatively,** incubation at 60°C for 2 min may improve the recovery for DNA larger than 3 kb.
9. **Remove beads** by using magnet rack, pipette DNA out and transfer to a clean tube.
10. **Store purified** DNA at -20°C for long-term storage.



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