

# NEUROMICS



## E18 Primary Mouse Hippocampal Cells

---

**Catalog Number:** PC35113

**Components :** •4 Cortical Pair Cord (2 mls)

Approximately  $1 \times 10^6$  cells/Cortical Pair

•12 mls Culture Media- Neurobasal/B27/0.5 mM glutamine/25  $\mu$ M glutamate culture medium; 5 mls Hibernat-Ca

**Description:** C57 Primary Mouse Hippocampal Cells are live neurons isolated from micro-surgically dissected regions of day 18 embryonic Mouse brain. These cells are prepared fresh each week and shipped in a nutrient rich medium that keeps the cells alive for up to 14 days under refrigeration.

**Shipping/Storage:** Primary Mouse Hippocampal Cells are shipped refrigerated. Cells are stable for up to 2 weeks when stored at 4-8°C. It is recommended to plate the cells as soon as possible after receiving cells according to Application Notes outlined below.

---

### Application Notes

#### Materials Needed Not Provide

- Poly-D-lysine (Sigma P6407) for substrate
- Papain (Sigma P4762; or Worthington) for enzymatic dissociation
- Trypan blue to count cells to get proper plating density
- Sterile pipette tips or sterile Pasteur pipette
- Sterile centrifuge tubes
- Centrifuge to operate at 200xg
- Water bath at 30°C
- General cell culture supplies (culture plates, coverslips, etc.)
- Additional media
  - Neurobasal (Invitrogen 1103-049)
  - B27 (Invitrogen 17504-044)
  - Glutamine (Invitrogen 35050-061)

#### Substrate Preparation

1. Prepare culture plate by coating with poly-D-lysine (0.15 ml/cm<sup>2</sup>, 50  $\mu$ g/ml, 135 kD) 1-20 hr., and rinse one time with 18 Mohm deionized water, and let dry.

#### Preparation of Isolated Neurons

1. Store tissue at 4°C until ready to use.
2. When ready to plate, make up 2 mls of enzymatic solution in shipping media without B27 (Hibernat-Ca; 5 mls supplied) containing 4 mgs (2mgs/ml) of papain. Make sure to sterile filter solution with a 0.2 micron filter after adding papain if source of enzyme is not sterile
3. Transfer the 2 ml of medium from the tissue tube into a 15 ml screw cap sterile tube; be careful not to disturb or remove tissue from the original tube. Save, do not discard.
4. Add 2 ml of media made in step 2 (Hibernat-Ca containing 2mgs/ml of papain). Incubate for 30 minutes at 30°C.
5. Remove enzymatic solution, again careful not to disturb or remove tissue, add back 1 ml of media saved in 15 ml tube.

---

FOR RESEARCH USE ONLY V4-01/08

Neuromics' reagents are for in vitro and certain non-human in vivo experimental use only and not intended for use in any human clinical investigation, diagnosis, prognosis, or treatment. We disclaim all liability in connection with the use of the information contained herein or otherwise, and all such risks are assumed by the user.

---

#### Preparation of Isolated Neurons-continued

6. Using a 1 ml pipettor with a sterile blue plastic tip, or a silanized 9-inch Pasteur pipette with the tip barely fire polished (preferable), suck the tissue with the medium into the pipette and immediately dispense the contents back into the same container. Take care not to create bubbles. Repeat this titration step about 10 times or until most all the cells are dispersed.
7. Let undispersed pieces settle by gravity for 1 min.
8. Transfer the dispersed cells (supernatant) into the 15 ml tube that contains the 1ml of media from Step 2, and gently mix the cells by swirling.
9. Spin the cells at 1,100 rpm (200xg) for 1 minute. Discard the supernatant while being careful not to remove any of the cells from the cell pellet.
10. Flick the tube a few times to loosen the cell pellet. Resuspend pellet in 1 ml of the provided B27/Neurobasal/0.5 mM glutamine medium. Resuspend cells by gently pipetting up and down. For E18 Hippocampus, media includes 25uM glutamate.
11. Aliquot 20  $\mu$ l and mix with 20  $\mu$ l of 0.4% trypan blue.
12. Count cells with a hemocytometer.
13. Further dilute the cells with B27/Neurobasal/0.5 mM glutamine to the desired plating density. We recommend  $32 \times 10^3$  cells/ $2 \text{ cm}^2$  of substrate in 0.4 ml/ $2 \text{ cm}^2$  substrate.
14. Incubate the cells at 37°C with 5% CO<sub>2</sub> and/or 9% or 20% oxygen.
15. After 4 days or longer, neurons are well differentiated. If further culture is desired, change half of medium with fresh, warm B27/Neurobasal/0.5 mM glutamine, without glutamate. Change half the medium every 3-4 days. Additional media and media supplements will need to be purchased to culture neurons past 4-6 days.

---

FOR RESEARCH USE ONLY V4-01/08

Neuromics' reagents are for in vitro and certain non-human in vivo experimental use only and not intended for use in any human clinical investigation, diagnosis, prognosis, or treatment. We disclaim all liability in connection with the use of the information contained herein or otherwise, and all such risks are assumed by the user.