

# Human Bladder Microvascular Endothelial Cells (HBMECs)

Catalog #: HEC21-T25 Cell #: > 90% confluent at (5X10<sup>5</sup> cells) in T25 flask

**Storage:** 37°C CO<sub>2</sub> incubator **Product Format:** Proliferating culture

#### **General Information**

HBMECs were initiated by elutriation from dissociated normal human bladder tissue. Passage 1 cells are ship in proliferating culture with a confluency of >90 %. ENDO-Growth Medium (MED001) is recommended for culture. Cells have an average additional population doubling levels >15 when cultured.

#### Characterization of the cells

- Cytoplasmic VWF / Factor VIII: >95% positive by immunofluorescence
- Cytoplasmic uptake of Di-I-Ac-LDL: >95% positive by immunofluorescence
- Cytoplasmic PECAM1: >95% positive by immunofluorescence

HBMECs are negative for HIV-1, HBV, HCV, and mycoplasma.

## **Recommended Products**

- ENDO-Growth Media MED001
  - Contains 475 ml of ENDO-Basal Media and 25 ml of ENDO-Growth Supplement combined. Which is freshly prepared for your convenience

OR

- ENDO-Growth Kit EGK001
  - Contains 475 of ENDO-Basal Media and 25 ml of ENDO-Growth Supplement in separately to be mixed to make growth media
- Smooth Coat Solution SC300
  - Biocompatible complex of extracellular matrix binding solution

OR

- AlphaBioCoat Solution AC001
  - Premium Smooth Coat Solution. Biocompatible complex of extracellular matrix binding solution with growth factors. Ideal for culturing cells from frozen.
- Cell Detachment Solution ADF001
  - Contains protease and collagenase activities in an isotonic, phosphate buffer solution with EDTA to detach primary cells and cell lines
- 1X Phosphate Buffer Solution PBS300

#### **Shipping**

Proliferating culture in T25 flask.

## **Handling of Arriving Cells**

When you receive the cells, leave the flask in 37°C CO2 incubator for 1 hour first, and then replace the transport medium with fresh Full medium. Let the cells grow for 24 hours before subculture.

# FOR RESEARCH USE ONLY

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Note: Handling human derived products is potentially biohazardous. Although each cell strain tests negative for HIV, HBV and HCV DNA, diagnostic tests are not necessarily 100% accurate, therefore, proper precautions must be taken to avoid inadvertent exposure. Always wear gloves and safety glasses when working these materials. Never mouth pipette. We recommend following the universal procedures for handling products of human origin as the minimum precaution against contamination.

# SUBCULTURE PROTOCOL (Proliferating in Culture)

**Note:** If you have any questions or need clarification regarding the protocol for culturing these cells, please reach out to Dr. Jensen Auguste at (978) 608-1766 with your questions before beginning.

- Coating T25 flasks:
  - Add 2 ml AlphaBioCoat Solution (AC001) into a T25 flask and ensure entire interior surface is coated with solution. After 30 minutes, dispose of Smooth Coat Solution by aspiration. Gently rinse and aspirate flask with phosphate buffer solution (PBS300). The flask is now ready for use(no need for overnight incubation when coated with AC001)
  - b. If you are using the coated flask the same day, add about 4 ml of Endo-Growth media (MED001) to the coated flask. \*If the media changes color from pink to yellow, aspirate and discard the media. Add 4ml of fresh media to the coated flask.
- Inspect to the confluence of the flask. If the flask is not 90% confluence, remove transport media and add 5ml of fresh media to the flask. Place flask in 37°C incubator until cells are at 90% confluence. Change media every 2 days.
- 3. If flask is at 90% confluence, aspirate transport media from flask
- Rinse T25 flask containing cells with 5 ml 1XPBS (PBS300).
- 5. Gently aspirate out the PBS after rinsing, and discard.
- Add 2ml of RT trypsin/ EDTA or Cell Detachement Solution (ADF001) to T25 flask containing cells (ensure entire interior surface is cover).
- 7. Place T25 flask containing cells into 37°C incubator for 1 or 2 minutes (cells will normally come off of the surface within 1 or 2 minutes).
- 8. Suspend the cells with 15ml of ENDO-Growth medium (MED001) and transfer equally into 3 pre-coated T25 flasks (the cells are now at a subculture ratio of 1:3).
- 9. There is no need to spin cells during subculture.
- 10. Proliferating cell culture: ENDO-Growth medium (MED001) should be changed every 2 days. The cells normally become confluent within 7 days ( when split at a 1:3 ratio)
- 11. Use ENDO- Basal media (MED002) containing 0.5% FBS to induce quiescent cells (after 18-24 hours)

Note: Should any issues arise while using our cells, our team is here to help troubleshoot any issues. Our cells are backed by our one-time replacement or refund policy. Our recommended protocol including recommended products must be used to be eligible for replacement or refund. Cells that have been refrozen are no longer eligible for refund or replacement.

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