NEUROMICS

Insulin Secreting Human Fibroblast Pancreatic Islet Cells

Catalog #: HPF002 **Cell #:** >5x10^5 cells

Storage: Liquid Nitrogen vapor phase until ready for culture. **Product Format:** Frozen Vial
While culturing keep in 37°C CO_2 incubator (95% air, 5% CO_2)

GENERAL INFORMATION

Insulin Secreting Human Fibroblast Pancreatic Islet Cells derived from a Human Pancreatic Islet Cells transformed using a proprietary method. The cell line can be expanded using our Stem Cell Complete Media (cat. CAFM02). Insulin is detected in the culture supernatant at (20-30pmol). Cells are supplied in frozen vials with more than 5 x 10[5] cell/vial. Stem Cell Complete Media (cat. CAFM02) is highly recommended to culture the cells. Cells are guaranteed to be passaged 5 times at a 1 to 3 split ratio.

Product is for Research use only.

Frozen Vials are shipped in a Dry Ice Package.

STATEMENT

Handling human tissue derived products is potentially bio-hazardous, despite testing negative for HIV, HBV, and HCV DNA. Nonetheless, proper precautions must be taken to avoid inadvertent exposure.

SPECIAL NOTES:

- We strongly advise our customers to use medium and related products recommended by Neuromics, because our cells were grown and adapted using our products.
- The growth featured of our cells cannot be guaranteed if the specific growth mediums stated in our datasheets are not used.
- Due to the sensitive nature of primary cells and cell lines, all quality related issues about the cells must be reported back to us within one month after receiving the products. Cells will not be replaced after the one-month period.

UNPACKING AND STORAGE INSTRUCTIONS

1. Check all containers for leakage or breakage. Remove the frozen cells from the dry ice packaging and immediately place the cells at a temperature below -130°C, preferably in liquid nitrogen vapor, until ready for use.

2. Complete medium: The base medium for this cell line is Stem Cell Complete Media (cat. CAFM02).

www.neuromics.com

Neuromics Antibodies • 5325 West 74th Street, Suite 8 • Edina, MN 55439
phone 866-350-1500 • fax 612-677-3976 • e-mail pshuster@neuromics.com

HANDLING PROCEDURE

To ensure the highest level of viability, thaw the vial and initiate the culture as soon as possible upon receipt. If upon arrival, continued storage of the frozen culture is necessary, it should be stored in liquid nitrogen vapor phase and not at -70°C. Storage at -70°C will result in loss of viability. Thaw the vial by gentle agitation in a 37°C water bath. To reduce the possibility of contamination, keep the O-ring and cap out of the water. Thawing should be rapid (approximately 2 minutes).

PROTOCOL FOR THAWING THE CELLS AND SUBCULTURE

Note: If you have any questions or need clarification regarding the protocol for culturing these cells, please reach out to Dr. Jensen Auguste at (978) 608-1766 with your questions before beginning.

1. Pre-coating of T25 flasks - Add 3-5ml AlphaBioCoat Solution (cat. AC001) into a T25 flask to cover the whole surface of the flask, 5 mins later, aspirate off the excessive coating solution and rinse the flask 1xPBS the flask is ready to be used.

2. Thaw the frozen cell vial in a 37C water bath first, and then transfer the cells into the pre-coated T25 flask with 15ml of growth media, cells usually become confluent within 5-7 days.

3. To passage the cells, rinse the cells in the T25 flask with 15ml HBSS (RT) twice; then add 2ml Cell Detachment solution (cat. ADF001) into one T25 flask; gently disposing excessive solution within 20 seconds by aspiration.

4. Leave the flask with the cells at RT or 37C for 1 min (most cells usually will detach from the surface within 1-2 mins monitor the cells under a microscope until most of the cells become rounded up, and then gently tap the flask against the bench surface, and the cells will move on the surface of the flask when monitoring under a microscope.

5. Add 5ml Neutralization Buffer (cat. NB001) and spin down the cells with 800g centrifugation for 5 mins.

6. Resuspend the cell pellet with 10 or 15ml growth media and transfer 5 ml each into 2 or 3 pre-coated T25 flasks (for 1/2 to 1/3 subculture ratio).

7. Change medium every 2 or 3days and the cells usually become confluent within 7 days (when split at a 1/3 ratio).

Subcultivation Ratio: A subcultivation ratio of 1:2 to 1:3 is recommended.

Medium Renewal: Every 2 to 3 days

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