NEUROMICS

GFP Expressing Human Brain Glioblastoma Cells (LN-18)

Catalog #: TR05-GFP

Cell #: >5x10^5 cells

Storage: Liquid Nitrogen until ready for culture.
While Culturing keep in 37°C CO_2 incubator

Product Format: Frozen Vial

GENERAL INFORMATION

Human Brain Glioblastoma Cells (LN-18) were derived from a Human Brain Glioblastoma of a 65-year old Caucasian male. GFP-Human Brain Glioblastoma Cells (LN-18) are selected from the Human Brain Glioblastoma Cells (LN-18) and transfected with GFP expressing lentiviruses resistant to puromycin. Cells are supplied in frozen vials with more than 5 x 10^5 cell/vial. Universal Full Growth Medium (TM001) is recommended to culture the cells.

Product is for Research use only.

Frozen Vials are shipped in a Dry Ice Package.

CHARACTERIZATION OF THE CELLS

Human Brain Glioblastoma cells (LN-18) are tested negative for HIV-1, HBV, HCV, and mycoplasma.

HANDLING OF ARRIVING CELLS

When you receive the dry ice package with cells in frozen vials, transfer the frozen vials of cells into a -80ºC freezer for short period storage or a liquid nitrogen tank for long-term storage.

PROTOCOL FOR THAWING THE CELLS AND SUBCULTURE

Note: If you have any questions or need clarification regarding the protocol for culturing these cells, please reach out to Dr. Jensen Auguste at (978) 608-1766 with your questions before beginning.

1. Pre-coating of T25 flasks-Add 2 ml each Universal Coating Solution (AC002) into a T25 flask to cover the whole surface of the flask, 5 mins later, dispose the excessive coating solution by aspiration and the flask is ready to be used (although solution containing other extracellular matrix, i.e. gelatin, collagen, and fibronectin, can be used, make sure to optimize the conditions in advance.

2. Thaw the frozen cell vial in a 37ºC water bath first, and then transfer the cells into the pre-coated T25 flask with 10 ml of Universal Full Growth Medium (TM001), cells usually become confluent with 5-7 days.

3. To passage the cells, rinse the cells in a T25 flask with 5 ml HBSS (Room Temperature) twice; then add 2 ml Universal Detachment Solution (RT) (AD002) into one T25 flask; gently dispose the excessive Universal Detachment Solution within 20 seconds by aspiration.

4. Leave the T25 flask with the cells at RT or 37ºC for 1 min (most cells usually will detach from the surface within 1-2 mins) or monitor the cells under a microscope until most of cells become rounded up, and then gently tap the flask against the bench surface, and the cells will move on the surface of the flask when monitoring under microscope.

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5. Add 5 ml of Universal Neutralization Buffer (NB001) and spin down the cells with an 800 g centrifugation for 5 mins.

6. Re-suspend the cell pellet with 10 ml or 15 ml of Universal Full Growth Medium and transfer 5 ml each into 2 or 3 pre-coated T25 flasks (for 1/2 to 1/3 subculture ratio).

7. Change the medium every 2 or 3 days and the cells usually become confluent within 7 days (when split at a 1/3 ratio).

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