

# NEUROMICS

## ACE2-GFP Human Pulmonary Bronchial/Tracheal Epithelial Cells (HPuBTEPCs-ACE2-GFP)

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**Catalog #:** HPEC003

**Cell #:** > 5X10<sup>5</sup> cells

**Storage:** Liquid Nitrogen until ready for culture.  
While culturing keep in 37°C CO<sub>2</sub> incubator

**Product Format:** Frozen Vial

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### GENERAL INFORMATION

Human Pulmonary Bronchial/Tracheal Epithelial Cells were isolated from normal human lung bronchial/tracheal tissue. HPuBTEPCs-ACE2-GFP were selected from puromycin resistant HPuBTEPCs after infected with lentiviruses expressing ACE2-GFP. The cells are shipped in frozen vials (the cells are provided @ passage 1). Human Epithelial Cell Medium (ECM001) is recommended for cell culture and these cells have a minimum average population doubling levels > 8 when cultured following the detailed protocol described below).

*Product is for Research use only*

Frozen vials are shipped in a dry ice package

### CHARACTERIZATION OF THE CELLS

1. Pan Cytokeratin Positive
2. E-7 Negative
3. Typical cobblestone monolayer morphology in culture
4. HPuBTEPCs are negative for HIV-1, HBV, HCV, and mycoplasma

### HANDLING OF ARRIVING CELLS

When you receive the dry ice package with cells in frozen vials, transfer the frozen vials of cells into a -80°C freezer for short period storage or a liquid nitrogen tank for long- term storage.

### PROTOCOL FOR THAWING THE CELLS AND SUBCULTURE

**Note:** If you have any questions or need clarification regarding the protocol for culturing these cells, please reach out to Dr. Jensen Auguste at (978) 608-1766 with your questions before beginning.

1. Pre-coating of T25 flasks- Add 2ml of AlphaBio Coat solution into a T25 flask to cover the whole surface of the flask, 5 mins later, dispose the excessive coating solution by aspiration and the flask is ready to be used.
2. Thaw the frozen cell vial in a 37C water bath first, and then transfer the cells into the pre-coated T25 flask with 10ml of Human Epithelial Cell Medium (ECM001), cells usually become confluent with 1-2 days and ready to be passaged.
3. To passage the cells, rinse the cells in a T25 flask with 5ml HBSS (RT) twice; then add 2ml Universal Detachment solution (RT) (AD002) into one T25 flask; gently dispose the excessive detachment solution within 20 seconds by aspiration.
4. Leave the T25 flask with the cells at RT or 37C for 1 min (most cells usually will detach from the surface within 1-2 mins; or monitor the cells under a microscope until most of cells become rounded up, and then gently tap the flask against the bench surface, and the cells will move on the surface of the flask when monitoring under microscope.
5. Add 5ml Universal Neutralization Buffer (NB001) and spin down the cells with 800g centrifugation for 5 mins.

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Neuromics Antibodies • 5325 West 74<sup>th</sup> Street, Suite 8 • Edina, MN 55439  
Phone 866-350-1500 • fax 612-677-3976 • email: [pshuster@neuromics.com](mailto:pshuster@neuromics.com)

6. Re-suspend the cell pellet with 10 or 15 ml medium and transfer 5 ml each into 2 or 3 pre-coated T25 flasks (for 1/2 to 1/3 subculture ratio).
7. Change medium every 2 or 3 days and the cells usually become confluent within 7 days (when split at a 1/3 ratio).
8. To prepare quiescent cells, when cells are nearly confluent, replace with Human Epithelial Basal Cell Medium (ECM002) containing 0.5%FBS for about 8-12hrs before your experiments.

Note: Should any issues arise while using our cells, our team is here to help troubleshoot any issues. Our cells are backed by our one-time replacement or refund policy. Our recommended protocol including recommended products must be used to be eligible for replacement or refund. Cells that have been refrozen are no longer eligible for refund or replacement.

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