Human Prostate Carcinoma Cells (LNCaP – Clone FGC)

Catalog #: LNC001

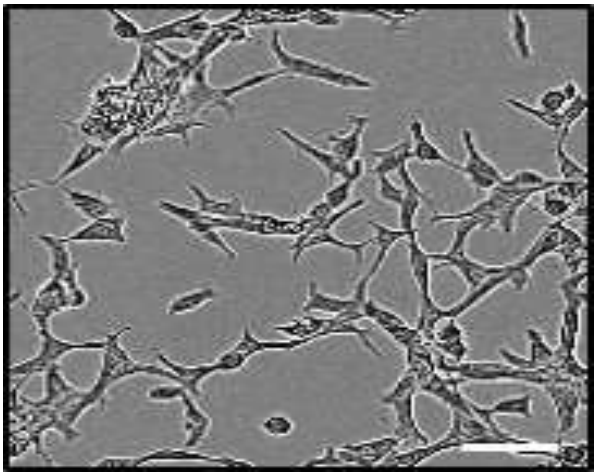
Cell #: >5x10^5 cells

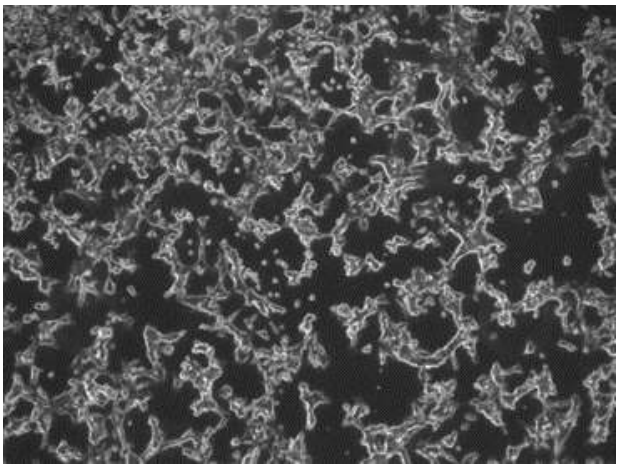
Storage: Liquid Nitrogen until ready for culture.
While Culturing keep in 37°C CO_2 incubator

Product Format: Frozen Vial

GENERAL INFORMATION

Human Prostate Carcinoma Cells (LNCaP Clone FGC) were derived from a prostate carcinoma patient. LNCaP clone FGC was isolated in 1977 by J.S. Horoszewicz, et al., from a needle aspiration biopsy of the left supraclavicular lymph node of a 50-year-old Caucasian male (blood type B+) with a confirmed diagnosis of metastatic prostate carcinoma. LNCaP cells are sensitive to androgens unlike other prostate cancer cell lines. Cells are supplied in frozen vials with more than 5x10^5 cells/vial. Universal Full Growth Medium (TM001) is recommended to culture the cells.





Product is for Research use only.

Frozen Vials are shipped in a Dry Ice Package.

CHARACTERIZATION OF THE CELLS

Human Prostate Carcinoma Cells (LNCaP Clone FGC) are tested negative for HIV-1, HBV, HCV, and mycoplasma.

HANDLING OF ARRIVING CELLS

To ensure the highest level of viability, thaw the vial and initiate the culture as soon as possible upon receipt. If additional storage is necessary, when you receive the dry ice package with cells in frozen vials, transfer the frozen vials of cells into a -80ºC freezer for short period storage or a liquid nitrogen tank for long-term storage.

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phone 866-350-1500 • fax 612-677-3976 • e-mail pshuster@neuromics.com

PROTOCOL FOR THAWING THE CELLS

1. Thaw the vial by gentle agitation in a 37°C water bath. To reduce the possibility of contamination, keep the O-ring and cap out of the water. Thawing should be rapid (approximately 2 minutes).

2. Remove the vial from the water bath as soon as the contents are thawed and decontaminate by dipping in or spraying with 70% ethanol. All operations from this point on should be carried out under strict aseptic conditions.

3. Transfer the vial contents to a 75 cm^2 tissue culture flask and dilute with the recommended complete culture medium (see the specific batch information for the recommended dilution ratio). It is important to avoid excessive alkalinity of the medium during recovery of the cells. It is suggested that, prior to the addition of the vial contents, the culture vessel containing the growth medium be placed into the incubator for at least 15 minutes to allow the medium to reach its normal pH (7.0 to 7.6).

4. Incubate the culture at 37°C in a suitable incubator. A 5% CO2 in air atmosphere is recommended if using the medium described on this product sheet.

Note: If it is desired that the cryoprotective agent be removed immediately, or that a more concentrated cell suspension be obtained, centrifuge the cell suspension at approximately 125 x g for 5 to 10 minutes. Discard the supernatant and resuspend the cells with fresh growth medium at the dilution ratio recommended in the specific batch information.

PROTOCOL FOR SUBCULTURE

Note: If you have any questions or need clarification regarding the protocol for culturing these cells, please reach out to Dr. Jensen Auguste at (978) 608-1766 with your questions before beginning.

Volumes are given for a 75 cm^2 flask. Increase or decrease the amount of dissociation medium needed proportionally for culture vessels of other sizes.

1. Remove and discard culture medium.

2. Briefly rinse the cell layer with Hank's Balanced Salt solution to remove all traces of serum that contains trypsin inhibitor.

3. Add 2.0 to 3.0 mL of Cell Detachment Solution (ADF001) to flask and observe cells under an inverted microscope until cell layer is dispersed (usually within 5 to 15 minutes). **Note:** To avoid clumping do not agitate the cells by hitting or shaking the flask while waiting for the cells to detach. Cells that are difficult to detach may be placed at 37°C to facilitate dispersal.

4. Add 6.0 to 8.0 mL of complete growth medium and aspirate cells by gently pipetting.

5. Add appropriate aliquots of the cell suspension to new culture vessels.

6. Incubate cultures at 37°C.

Subcultivation Ratio: A subcultivation ratio of 1:3 to 1:6 is recommended

Medium Renewal: 2 to 3 times per week

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