



RFP Expressing Human Gastric Carcinoma N87 Cells

Catalog #: TR02-RFP

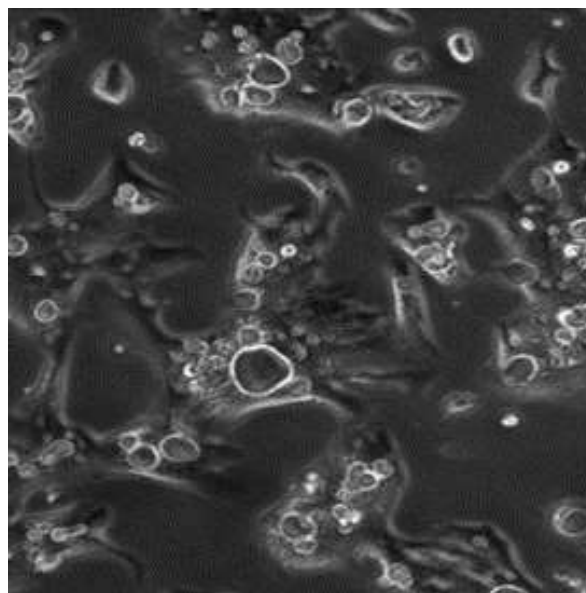
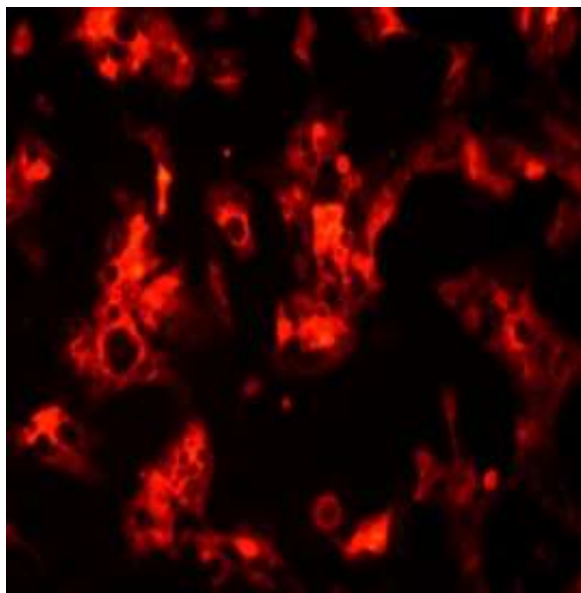
Cell #: >5x10⁵ cells

Storage: Liquid Nitrogen until ready for culture.
While Culturing keep in 37°C CO₂ incubator

Product Format: Frozen Vial

GENERAL INFORMATION

Human Gastric Carcinoma N87 Cells (N87 cells) were established from a gastric carcinoma patient. RFP-Tagged Human Gastric Carcinoma N87 Cells (RFP-N87 cells) are selected from N87 cells after infected with lentiviruses expressing RFP with puromycin. The cells can be cultivated with Universal Full Growth Medium (TM001) and they have a minimum population doubling capacity > 12 when cultured following the detailed protocol described below.



Product is for Research use only.

Frozen Vials are shipped in a Dry Ice Package.

CHARACTERIZATION OF THE CELLS

N87 cells are negative for HIV-1, HBV, HCV, and mycoplasma.

HANDLING OF ARRIVING CELLS

When you receive the dry ice package with cells in frozen vials, transfer the frozen vials of cells into a -80°C freezer for short period storage or a liquid nitrogen tank for long-term storage.

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PROTOCOL FOR THAWING THE CELLS AND SUBCULTURE

Note: If you have any questions or need clarification regarding the protocol for culturing these cells, please reach out to Dr. Jensen Auguste at (978) 608-1766 with your questions before beginning.

1. Pre-coating of T25 flasks: Add 1 ml of Fibronectin (FB001) into one T25 flask and make sure whole surface of the flask is covered with the coating solution and leave flask at room temperature for overnight or 37°C for 1 hour. Dispose excessive fibronectin by aspiration. Rinsing the flask with PBS once before the flask is ready to be used.
2. Thaw the frozen cell vial in a 37°C water bath first, and then transfer the cells into the pre-coated T25 flask with 10 ml of Universal Full Growth Medium (TM001), cells usually become confluent with 5-7 days.
3. To passage the cells, rinse the cells in a T25 flask with 5 ml HBSS (Room Temperature) twice; then add 2 ml Universal Detachment Solution (RT) (AD002) into one T25 flask; gently dispose the excessive Universal Detachment Solution within 20 seconds by aspiration.
4. Leave the T25 flask with the cells at RT or 37°C for 1 min (most cells usually will detach from the surface within 1-2 mins) or monitor the cells under a microscope until most of cells become rounded up, and then gently tap the flask against the bench surface, and the cells will move on the surface of the flask when monitoring under microscope.
5. Add 5 ml of Universal Neutralization Buffer (NB001) and spin down the cells with an 800 g centrifugation for 5 mins.
6. Re-suspend the cell pellet with 10 ml or 15 ml of Universal Full Growth Medium and transfer 5 ml each into 2 or 3 pre-coated T25 flasks (for 1/2 to 1/3 subculture ratio).
7. Change the medium every 2 or 3 days and the cells usually become confluent within 7 days (when split at a 1/3 ratio).

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